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Incidence and Antimicrobial Susceptibilty Pattern of Urinary Pathogens Among Patients Attending Sir Yahaya Memorial Hospital Birnin Kebbi, Kebbi State Nigeria

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ABSTRACT

This study investigated the incidence and antibiotic susceptibility pattern of urinary pathogens commonly isolated from patients suspected of UTI at Sir Yahaya Memorial Hospital Birnin Kebbi, Kebbi State Nigeria. Two hundred urine samples from patients suspected with urinary tract infection were collected, the causative organisms were isolated on CLED and blood agar and identified using macroscopy, microscopy and biochemical test and their antimicrobial susceptibility patterns were profiled using Kirby Bauer method. The overall incidence of UTI was found to be 31.5% (63/200). Staphylococcus aureus 31/200 (15.5%), Klebsiella spp 20/200 (10%) and Proteus spp 12/200 (6%). Staphylococcus aureus, showed high percentage of susceptibility to the antibiotics viz; ampicillin (58.0%), amoxicillin (88%), cotrimoxazole (93.5%) and pefloxacin (96.8%), resistant to ceftazidime and gentamicin (83.9%), and ofloxacin and streptomycin (100%). Klebsiella spp was susceptible to amoxicillin (95%) and (100%) to ciprofloxacin but resistant to the seven remaining antibiotics. *Proteus* spp showed (100%) resistance to ampicillin, ciprofloxacin, cotrimoxazole and streptomycin, but susceptible to amoxicillin, ofloxacin, and pefloxacin (50%). In conclusion, the three isolated uropathogens were susceptible to amoxicillin (> 80%), resistant to streptomycin, ofloxacin and gentamicin (<10%). Thus, routine surveillance and regular monitoring is required to establish reliable information about susceptibility pattern of urinary pathogens for optimal empirical therapy of patients with UTI.

Keywords: Multi antibiotic resistance, Prevalence, Urinary tract infections, Kebbi State

INTRODUCTION

Urinary tract Infection (UTI) is one of the most common microbial infection encountered in general practices (Flores-Mireles et al., 2015; Iwuafor et al., 2016; Cortes-Penfield et al., 2017). Both young and adults have a history of at least one episode of UTI in their lifetime (Dibua et. al., 2014; Khan et al., 2017). Community acquired infection is caused by Escherichia coli, Klebsiella pneumoniae, Proteus mirabilis, Staphylococcus saprophyticus or Enterococcus faecalis, while the hospital acquired infection were caused by Escherichia coli, Pseudomonas aeruginosa, Proteus sp, Enterobacter sp., Serratia sp. or Enterococcus (Hanna-Wakim et al. 2015; Bernabé et al. 2017; Khan et al., 2017). The incidence of both community acquired and nosocomial UTI among patients rises annually. A significant number of patients who develop UTI will have recurrent urinary tract infections due to antimicrobial resistance (Sievert et al., 2013; Flores-Mireles et al., 2015).

The prevalence of UTI and subsequent increasing antimicrobial resistance can vary according to geographical and regional location (Mordi et al., 2016; Oli et al., 2015). In Nigeria, the prevalence of community-acquired and nosocomial UTIs conducted in a hospital among 12458 urine samples were 12.3% and 9.3%, respectively (Iregbu and Nwajiobi-Princewill, 2013). In a related study conducted in Ile-Ife, Nigeria, the incidence of UTIs among children and adolescence was reported to be 11.9%, of which 22.4% were females while 4.6% were males ((Flores-Mireles et al., 2015; Iwuafor et al., 2016). Factors such as gender, age, urinary catheter, genitourinary tract abnormalities, pregnancy, infants, elderly, hospitalization status were attributed to high risk for recurrent UTIs (Hidron et al., 2008; Iregbu and Nwajiobi-Princewill, 2013; Oli et al., 2015). In addition, lack of hygiene, compliance to dose and duration of the prescribed antibiotic, unjustified prescription written by doctors; and easy access to over the counter medication makes the management of UTI a challenging task (Khan et al., 2017; Odoki et al., 2019).

Multi antibiotic resistant bacteria have been reported in Kebbi State (WHO, 2014). However, no documented literature on uropathogens and their antibiotic susceptibility pattern among patients attending various hospitals in the State. This study thus, was undertaken to find out the incidence and antibiotic susceptibility pattern of isolates obtain from urine of patients attending Sir Yahaya Memorial Hospital Birnin Kebbi (SYMH) Kebbi State. Knowledge of the uropathogens and their antimicrobial resistance profiles may assist in administering the appropriate antimicrobial treatment.

MATERIALS AND METHODS **Study Population**

The study was conducted for a period of 10 months from January 2018 to October 2018 among UTI patients attending SYMH Birnin Kebbi, in Kebbi State, Northwest Region of Nigeria. The study included 200 patients with suspected cases of urinary tract infection. Information on patient's age and gender was obtained from the Laboratory Information System. Ethical clearance was obtained prior to data collection from the Kebbi State Ministry of Health Ethics and Research Committee, Gwadangwaji Secretariat, Birnin Kebbi, Nigeria. The study subjects were enrolled

after verbal informed consent was obtained. In case of minors, consents were sought from the parent or guardian.

Sample Collection and Processing

Clean urine samples were collected in sterile urine collection tubes. The urine samples were examined macroscopically and microscopically. Thereafter, $10\,\mu l$ of urine was inoculated using calibrated loops by a surface streak procedure onto both CLED and blood agar, and incubated at 37°C for 24 hours. A specimen was considered positive for UTI if a single organism was observed after 24 hours of incubation and negative after further incubation for 48 hours. UTI isolates obtained on the two media were identified following standard microscopic and biochemical procedures, namely indole, citrate, oxidase, H₂S production, lysine decarboxylase, lactose fermentation, urea hydrolysis, gas production, catalase, coagulase, manitol fermentation (Clarck and Cowan, 2009).

Susceptibility Testing

The antimicrobial susceptibility of the isolates to nine commonly prescribed antibiotics was determined using the Kirby-Bauer disk diffusion method (Bauer et al., 1978). A lawn of test pathogen (18 hours culture) from the nutrient agar was transferred into 5mls of sterile peptone broth. The suspension was adjusted to equal 0.5 McFarland turbidity standard. Subsequently, 3-4 drops of inoculum was spread across an entire surface of Mueller Hinton agar plate. The plates were allowed to dry. Then, antibiotic discs were gently and firmly placed on the agar plates, which were then left at room temperature for 1 hour to allow diffusion of the

antibiotics into the agar medium. The plates were then incubated at 37°C for 24 hours. After overnight incubation, growth inhibition was shown as a circular zone of no growth around the disc. The zones of growth inhibition around each of the antibiotic disks are measured to the nearest millimeter. The diameter of the zone is related to the susceptibility of the isolate and to the diffusion rate of the drug through the agar medium. The zone diameters of each antibiotic are interpreted using the criteria published by Clinical and Laboratory Standards Institute (CLSI) guidelines. The antibiotics used were Ampicillin (AM, 10 g), Amoxicillin (AX, 30 g), Ceftazidime (CAZ, 30 g), Ciprofloxacin (CIP, 5 g), Cotrimoxazole (COT, 30 g), Gentamicin (GEN, 10 g), Ofloxacin (OF, 5 g), Pefloxacin (PEF, 10 g), and Streptomycin (S, 10 g). Data were entered and analyzed using Excel 2013. Discrete variables were expressed as percentages.

RESULT

Out of the 200 urine samples, 105 samples had positive bacterial growth (52.5%), while 95 (47.5%) samples yielded no growth (Table 1). The highest growth was recorded in female patients 66 (33.0%) compared to males 33 (19.5%). Female patients within the age group of 0-15 and 16-30 years had the highest bacterial growth 31 (15.5%) and 20 (10.0%) respectively. The highest bacterial growth recorded in male patients was observed within the age group 0-15 years 15 (7.5%). Male patients within the age group of 31-45 years had the lowest bacterial growth 4 (2.0%), (Table

Table 1. Frequency of bacteria isolated from UTI suspected patients at SYMH Birnin Kebbi

	No Positive (n=105)		No Negative (n=95)		
Age distribution	Male	Female	Male	Female	
0 - 15	15(7.5)	31(15.5)	9(4.5)	10(5.0)	
16 -30	11(5.5)	20(10.0)	10(5.0)	19(9.5)	
31 - 45	4(2.0)	7(3.5)	10(5.0)	23(11.5)	
46 - 60	9(4.5)	8(4.0)	6(3.0)	8(4.0)	
Total	39(19.5)	66 (33)	35(17.5)	60 (30)	

The most frequently isolated uropathogens out of three was Staphylococcus aureus 31 (15.5%), followed by Klebsiella spp 20 (10.0%) and *Proteus* spp 12 (6.0%) (Table 2). Twenty one of the Staphylococcus aureus (10.5%) were isolated from male patients, and 12 (6.0%) of the isolates were from patients between the ages of 16 - 30 years and 17 (8.5%) were outpatients. Eleven of the Klebsiella spp

(5.5%) were from female patients, and 8 (4.0%) were obtained from patients aged 16 -30 years. Nine of the Proteus spp (4.5%) was isolated from male patients, 4 (2.0%) from patients aged 16-60 years and 7(3.5%) of the isolates were from outpatients (Table 2).

Table 2:	Age-related	incidence	of bacte	erial isolates	amonost	study subjects

Distribution	Staphylococcus aureus n=31(15.5%)	Klebsiella spp	Proteus spp	
		n=20 (10%)	n=12(6%)	
Male	21(10.5)	9(4.5)	9(4.5)	
Female	10(5.0)	11(5.5)	3(1.5)	
0 - 15	7(3.5)	5(2.5)	2(1.0)	
16 -30	12(6.0)	8(4.0)	3(1.5)	
31 - 45	7(3.5)	3(1.5)	4(2.0)	
46 - 60	5(2.5)	4(2.0)	3(1.5)	
Out patients	17(8.5)	1(0.5)	7(3.5)	
Inpatients	14(7.0)	19(9.5)	5(2.5)	

The three uropathogens isolated from patients with UTIs showed high levels of susceptibility to commonly prescribed antimicrobial drugs. Figure 1, shows percentage distribution of uropathogens susceptible to more than one of the nine commonly prescribed antibiotic at SYMH. From figure 1 below, 10 (32.3%) of Staphylococcus aureus were susceptible to more than 2 antibiotics, 20 (64.5%) susceptible to more than 5 antibiotics and 1 (3.2%) was susceptible to more than 7 antibiotics. Klebsiella spp 18 (90.0%) were susceptible to more than two antibiotics, 2 (10.0%) susceptible to more than 4 antibiotics. *Proteus* spp 10 (83.3%) and 2 (16.7%) were susceptible to more 2 and 5 antibiotics respectively, (Figure 1). None of the Klebsiella and proteus spp were susceptible to more than 6 antibiotics.

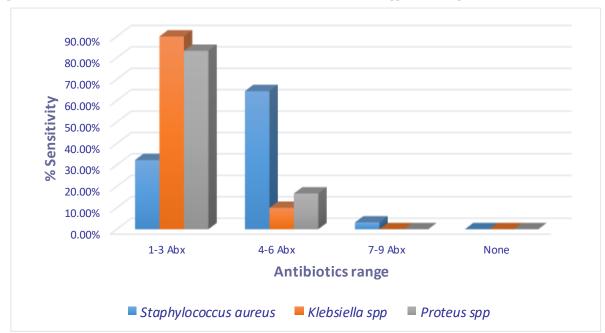


Figure 1: Number (%) of uropathogens susceptible to more than one of the test antibiotic

Table 3 shows the antibiotic susceptibility pattern of the uropathogen isolated. Staphylococcus aureus showed (58.0%) susceptibility to ampicillin, (88%) of the isolates were susceptible to amoxicillin, (93.5%) to cotrimoxazole and (96.8%) to pefloxacin. The organism was resistant to ceftazidime, gentamicin, ofloxacin and streptomycin. Klebsiella spp, which is the second most dominant uropathogen isolated, is (95%) susceptible to amoxicillin and (100%) to ciprofloxacin but resistant to the seven remaining antibiotics, (Table 3). Proteus spp showed

(100%)resistance ampicillin, ciprofloxacin, to cotrimoxazole and streptomycin, but susceptible to amoxicillin, ofloxacin, and pefloxacin (50%). An average of 80% of the three isolated uropathogens was susceptible to amoxicillin and more than 90% were resistant to streptomycin, ofloxacin and gentamicin (Table 3).

	- ·	Staphylococcus aureus Klebsiella spp (n=31) (n=20)		* *	Proteus spp (n=12)	
Antibiotics	Susceptible	Resistant	Susceptible	Resistant	Susceptible	Resistant
Ampicillin	18(58)	13(42)	0(0)	20(100)	0(0)	12(100)
Amoxicillin	27(88)	4(12)	19(95)	1(5)	6(50)	6(50)
Ceftazidime	5(16.1)	26(83.9)	4(20)	16(80)	1(8.3)	11(91.7)
Ciprofloxacin	1(3.2)	30(96.8)	20(100)	0(0)	0(0)	12(100)
Cotrimoxazole	29(93.5)	2(6.5)	0(0)	20(100)	0(0)	12(100)
Gentamicin	5(16.1)	26(83.9)	4(20)	16(80)	5(41.7)	7(59.3)
Ofloxacin	0(0)	31(100)	4(20)	16(80)	6(50)	6(50)
Pefloxacin	30(96.8)	1(3.2)	5(25)	15(75)	6(50)	6(50)
Streptomycin	0(0)	31(100)	2(10)	18(90)	0(0)	12(100)

Table 3: Susceptibility pattern of uropathogens isolated in urine culture at SYMH Birnin Kebbi from January – October 2018

DISCUSSION

Urinary tract infection caused by bacteria is one of the most common clinical cases that outpatients and inpatients seek treatment at SYMH Birnin Kebbi. Effective management of these patients depends on the identification of the causative agents and administering an effective antibiotic regimen. In this study, the isolation rate of uropathogens at Sir Yahaya Memorial Hospital Birnin Kebbi between the months of January 2018 to October 2018 from urine sample was 31.5%. This is comparatively lower than reports from previous studies in Okada community of Edo State (39.7%) (Henry Oladeinde et al., 2011), at Specialist Hospital Yola (67.2%) (Abubakar, 2009), Dalhatu Araf Specialist Hospital, Lafia, Nasarawa (60%) (Durowade et al., 2009), and among pregnant women in Ebonyi State (77.9%) (Onoh et al., 2013). The lower rate of isolation might have been due to sample size variation. However, this finding is in line with studies done at National Hospital in Abuja (13.1%) (Iregbu and Nwajiobi-Princewill, 2013), among antenatal patients in Nnewi (18.2%) (Oli et al., 2010) and amongst pregnant women in Amassoma, Southern Nigeria (25.3%) (Lawani et al., 2015).

The bacterial growth yields from urine samples were markedly different between males and females (male = 33.0% and female = 66.0%), and differ among age groups. This agrees with previous reports which shows that uropathogens were more frequent in females than males, because of close proximity of the female urethra to the gastrointestinal outlet making it easier for uropathogens to colonize this area (Iregbu and Nwajiobi-Princewill, 2013; Cortes-Penfield et al., 2017; Odoki et al., 2019). The most frequent and dominant uropathogens isolated from urine samples in both outpatients and inpatients of both sexes was Staphylococcus aureus, which made up 50 % of all the cases. This findings agrees with previous reports undertaken on asymptomatic subjects in an urban settlement in Benin City of Nigeria and on pregnant women in Oyo and Ebonyi State and on febrile children in Nasarawa State (Musa-Aisien et al., 2003; Okonko et al., 2009; Chukwudozie Onuoha, 2014). The very high rate of Staphylococcus aureus recorded in this study likely represent poor handling of the samples (Nicole, 2002; Foxman, 2003).

Klebsiella spp is the second most dominant uropathogen isolated in the study. The specie has the ability to spread rapidly in the hospital environment and cause nosocomial infection and outbreaks (Khan et al., 2017; Odoki et al.

2019;) because the reservoirs for transmission of *Klebsiella* spp are the gastrointestinal tract and the hands of hospital personnel (Podschun and Ullmann, 1998; Hanna-Wakim et al., 2015). The findings corroborate previous results as reported in two primary health care centers in Oshogbo, Osun State (Olawale et al., 2011), among pregnant women in Ondo State (Onoh et al., 2013), and among symptomatic patients attending tertiary hospitals in Abuja and Maiduguri in Borno State (Iregbu and Nwajiobi-Princewill, 2013; Anejo-okopi et al. 2015). Proteus spp were the least uropathogens isolated in this study. The proteus spp are very important uropathogens because of their ability to cause infections and are resistant to many antibiotics (Aiyegoro et al. 2007; Akingbade et al. 2014; Dibua et al. 2014; Bernabé et al. 2017). These findings agrees with previous studies (Kehinde et al. 2012; Mava et al. 2012; Iregbu and Nwajiobi-Princewill 2013; Chukwudozie Onuoha, 2014; Flores-Mireles et al. 2015).

This study revealed that a higher rate of susceptibility to the commonly prescribed antibiotic agent. The susceptibility profile indicates that the amoxicillin and cotrimoxazole was the most active antibacterial agent followed by the ciprofloxacin. This result is in agreement with recommendations by Hsueh et al. (2011); Iregbu and Nwajiobi-Princewill (2013); Olorunmola et al. (2013); Bernabé et al. (2017). These antibiotics could be considered as alternative options in the empirical treatment of UTIs. Streptomycin, ofloxacin and ceftazidime showed very poor activity. This may be due to long-term use of these drugs over the years without prescription (Flores-Mireles et al. 2015; Khan et al. 2017; Odoki et al. 2019). This implies that these antibiotics cannot be used as empirical therapy for urinary tract infection particularly in the study area. The variations in susceptibility of the three uropathogens to different antibiotics commonly prescribed may be due to the indiscriminate prescription of antibiotics (Hidron et al. 2008; Olorunmola et al. 2013; Sievert et al. 2013; Akingbade et al. 2014; Bernabé et al. 2017).

In conclusion, the most predominant uropathogen isolated in this study was Staphylococcus aureus followed by Klebsiella spp then Proteus spp. The overall incidence of UTI among patients attending Sir Yahaya Memorial Hospital Birnin Kebbi in Kebbi State was 31.5%. All the three uropathogens isolated were susceptible to amoxicillin (> 80%) and resistant to streptomycin, ofloxacin and

gentamicin (<10%). Therefore, there is need for routine surveillance and regular monitoring of susceptibility pattern of urinary pathogens among UTI patients attending Sir Yahaya Memorial Hospital Birnin Kebbi for optimal empirical therapy.

Author's contribution:

MBD, BA, AYB and ASK, conceived the study and participated in its design and coordination. MBD and ASK performed sample collection and analysis. MBD, BA, AYZ and ASK analyzed the results. MBD and ASK drafted the manuscript.

Conflict of interest:

All authors have no competing interests.

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REFERENCES

- Abubakar, E.-M. M. (2009). Antimicrobial susceptibility pattern of pathogenic bacteria causing urinary tract infections at the Specialist Hospital, Yola, Adamawa state, Nigeria. In Journal of Clinical Medicine and Research. 1(1) 001-008
- Anejo-okopi, A.I, Okwori A.E.J., Eze M.I., Onaji A.I., Ali M., Adekwu A, and Ejiji IS. (2015). Prevalence and Antibiotic Resistance Pattern of Urinary Tract Bacterial Infections Among Symptomatic Patients Attending University of Maiduguri Teaching Hospital, North East Nigeria. European Journal of Advanced Research in Biological and Life Sciences. 3;(3); 31-41
- Aiyegoro, A., Igbinosa, O., Ogunmwonyi, I., Odjadjare, E., Igbinosa, O., and Okoh, A. (2007). Incidence of urinary tract infections (UTI) among children and adolescents in Ile-Ife, Nigeria. African Journal of Microbiol Research. 013-019
- Akingbade, O., Balogun, S., Ojo, D., Akinduti, P., Okerentugba, P. O., Nwanze, J. C., and Okonko, I. O. (2014). Resistant plasmid profile analysis of multidrug resistant Escherichia coli isolated from urinary tract infections in Abeokuta, Nigeria. African Health Sciences. https://doi.org/10.4314/ahs.v14i4.8
- Bauer, A. W., Kirby, W. M. M., Sherris, J. C., Turck, A. M., and Graevenitz, A. Von. (1978). 40 Microbiology: A Centenary Perspective 1966 Antibiotic Susceptibility Testing by a Standardized Single Disk Method. American Journal of Clinical Pathology. https://doi.org/10.1016/S0305-4179(78)80006-0
- Bernabé, K. J., Langendorf, C., Ford, N., Ronat, J. B., and Murphy, R. A. (2017). Antimicrobial resistance in West Africa: a systematic review and meta-analysis. International Journal of Antimicrobial Agents. https://doi.org/10.1016/j.ijantimicag.2017.07.002
- Clarke, P. H., and Cowan S. T.. (2009). "Biochemical Methods for Bacteriology." Journal of General Microbiology. https://doi.org/10.1099/00221287-6-1-2-187.
- Chukwudozie Onuoha, S. (2014). Prevalence and Antimicrobial Susceptibility Pattern of Urinary Tract Infection(UTI) among Pregnant Women in Afikpo,

- Ebonyi State, Nigeria. American Journal of Life Sciences. https://doi.org/10.11648/j.ajls.20140202.12
- Cortes-Penfield, N. W., Trautner, B. W., and Jump, R. L. P. (2017). Urinary Tract Infection and Asymptomatic Bacteriuria in Older Adults. Infectious Disease Clinics North America. of https://doi.org/10.1016/j.idc.2017.07.002
- Dibua, U. M. E., Onyemerela, I. S., and Nweze, E. I. (2014). Frequency, Urinalysis and Susceptibility Profile of Pathogens Causing Urinary Track Infections in Enugu State, Southeast Nigeria. Revista Do Instituto de Medicina Tropical de São Paulo. https://doi.org/10.1590/s0036-46652014000100008
- Durowade, A. K., kolawole, A. S., Kolawole, O. M., Kandaki-Olukemi, Y. T., Babatunde, S. K., and Kolawole, C. F. (2009). Prevalence of urinary tract infections (UTI) among patients attending Dalhatu Araf Specialist Hospital, Lafia, Nasarawa State, Nigeria. International Journal of Medicine and Medical Sciences. 1 (5); 163-167
- Flores-Mireles, A. L., Walker, J. N., Caparon, M., and Hultgren, S. J. (2015). Urinary tract infections: Epidemiology, mechanisms of infection and treatment options. Nature Reviews Microbiology. https://doi.org/10.1038/nrmicro3432
- Foxman, B. (2003). Epidemiology of urinary tract infections: Incidence, morbidity, and economic costs. Disease-a-Month. https://doi.org/10.1016/S0011-5029(03)90000-9
- Hanna-Wakim, R. H., Ghanem, S. T., El Helou, M. W., Khafaja, S. A., Shaker, R. A., Hassan, S. A., and Dbaibo, G. S. (2015). Epidemiology and characteristics of urinary tract infections in children and adolescents. Frontiers in Cellular and Infection Microbiology. https://doi.org/10.3389/fcimb.2015.00045
- Henry Oladeinde, B., Omoregie, R., Olley, M., and Anunibe, J. A. (2011). Urinary tract infection in a rural community of Nigeria. North American Journal

Medical Sciences. https://doi.org/10.4297/najms.2011.375

- Hidron, A. I., Edwards, J. R., Patel, J., Horan, T. C., Sievert, D. M., Pollock, D. A., and Fridkin, S. K. (2008). Antimicrobial-Resistant Pathogens Associated With Healthcare-Associated Infections: Annual Summary of Data Reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2006-2007. Infection Control and Hospital Epidemiology. https://doi.org/10.1086/591861
- Hsueh, P. R., Hoban, D. J., Carmeli, Y., Chen, S. Y., Desikan, S., Alejandria, M., and Binh, T. Q. (2011). Consensus review of the epidemiology and appropriate antimicrobial therapy of complicated urinary tract infections in Asia-Pacific region. In Journal https://doi.org/10.1016/j.jinf.2011.05.015
- Iregbu, K., and Nwajiobi-Princewill, P. (2013). Urinary tract infections in a Tertiary Hospital in Abuja, Nigeria. In African Journal of Clinical and Experimental Microbiology. https://doi.org/10.4314/ ajcem. v14i3.9

- Iwuafor, A. A., Ogunsola, F. T., Oladele, R. O., Oduyebo, O. O., Desalu, I., Egwuatu, C. C., and Ogban, G. I. (2016). Incidence, clinical outcome and risk factors of intensive care unit infections in the lagos university teaching hospital (LUTH), Lagos, Nigeria. https://doi.org/10.1371/journal.pone.0165242
- Iregbu, K.C., and Nwajiobi-Princewill P.I. (2013). Urinary tract infections in a tertiary hospital in abuja, nigeria. African Journal of Clinical and Experimental Microbiology. Vol 14. No 3
- Kehinde, A., Adedapo, K., Aimakhu, C., Odukogbe, A. T., Olayemi, O., and Salako, B. (2012). Urinary pathogens and drug susceptibility patterns of urinary tract infections among antenatal clinic attendees in Ibadan, Nigeria. Journal of Obstetrics and Gynaecology Research. https://doi.org/10.1111/j.1447-0756.2011.01635.x
- Khan, H. A., Baig, F. K., and Mehboob, R. (2017). Nosocomial infections: Epidemiology, prevention, control and surveillance. Asian Pacific Journal of **Tropical** Biomedicine. https://doi.org/10.1016/j.apjtb.2017.01.019
- Lawani, E. U., Alade, T., and Oyelaran, D. (2015). Urinary tract infection amongst pregnant women in Amassoma, Southern Nigeria. African Journal of Microbiology Research. https://doi.org/10.5897/ajmr2014.7323
- Mava, Y., Bello, M., Ambe, J. P., and Zailani, S. B. (2012). Antimicrobial sensitivity pattern of organisms causing urinary tract infection in children with sickle cell anemia in Maiduguri, Nigeria. Nigerian Journal of Clinical Practice. https://doi.org/10.4103/1119-3077.104515
- Mordi, R. M., Burke, M. E., Odjadjare, E. E., Enabulele, S. A., and Umeh, O. J. (2016). Prevalence of Urinary Tract Infections (UTI) Among Pregnant Women in University Of Benin Teaching Hospital (UBTH) Benin City, Nigeria. Journal of Asian Scientific Research.
 - https://doi.org/10.18488/journal.2/2015.5.4/2.4.198.2
- Musa-Aisien, A. S., Ibadin, O. M., Ukoh, G., and Akpede, G. O. (2003). Prevalence and antimicrobial sensitivity pattern in urinary tract infection in febrile under-5s at a children's emergency unit in Nigeria. **Tropical** Annals of Paediatrics. https://doi.org/10.1179/000349803125002850
- Nicolle, L. E. (2002). Epidemiology of urinary tract infections. Clinical Microbiology Newsletter. https://doi.org/10.1016/S0196-4399(02)80035-6
- Odoki, M., Aliero, A. A., Tibyangye, J., Nyabayo Maniga, J., Wampande, E., Kato, C. D., and Bazira, J. (2019). Prevalence of Bacterial Urinary Tract Infections and Associated Factors among Patients Attending Hospitals in Bushenyi District, Uganda. International Journal Microbiology. https://doi.org/10.1155/2019/4246780
- Okonko, I. O., Ijandipe, L. A., Ilusanya, O. A., Donbraye-Emmanuel, O. B., Ejembi J., Udeze A. O., Egun O. C., Fowotade A. and Nkang A. O. (2009). Incidence of urinary tract infection (UTI) among pregnant

- women in Ibadan, South-Western Nigeria. Journal of Biotechnology. 8 (23); 6649-6657
- Olawale, K. O., Fadiora, S. O., and Taiwo, S. S. (2011). Prevalence of hospital-acquired enterococci infections in two primary-care hospitals in osogbo, southwestern Nigeria. African Journal of Infectious Diseases. 5(2): 40-46
- Oli, A., Ekejindu, C., Ejiofor, O., Oli, A., Ezeobi, I., and Ibeh, C. (2015). The Knowledge of and Attitude to Hospital-acquired Infections among Public and Private Healthcare Workers in South-East, Nigeria. British Journal of Medicine and Medical Research. https://doi.org/10.9734/bjmmr/2016/18272
- Oli, A. N., Okafor, C. I., Ibezim, E. C., Akujiobi, C. N., and Onwunzo, M. C. (2010). The prevalence and bacteriology of asymptomatic bacteriuria among antenatal patients in nnamdiazikiwe university teaching hospital nnewi; South Eastern Nigeria. Nigerian Journal of Clinical Practice. 13(4):409-12
- Olorunmola, F. O., Kolawole, D. O., and Lamikanra, A. (2013). Antibiotic resistance and virulence properties in Escherichia coli strains from cases of urinary tract infections. In African Journal of Infectious Diseases. 7(1): 1–7
- Onoh, R. C., Umeora, O. U. J., Egwuatu, V. E., Ezeonu, P. O., and Onoh, T. J. P. (2013). Antibiotic sensitivity pattern of uropathogens from pregnant women with urinary tract infection in Abakaliki, Nigeria. Infection Drug Resistance. https://doi.org/10.2147/IDR.S46002
- Podschun, R., and Ullmann, U. (1998). Klebsiella spp. as nosocomial pathogens: epidemiology, taxonomy, typing methods, and pathogenicity factors. Clinical Microbiology Reviews. 11(4):589-603
- Sievert, D. M., Ricks, P., Edwards, J. R., Schneider, A., Patel, J., Srinivasan, A., and Fridkin, S. (2013). Antimicrobial-Resistant Pathogens Associated with Healthcare-Associated Infections Summary of Data Reported to the National Healthcare Safety Network at the Centers for Disease Control and Prevention, 2009-2010. Infection Control and Hospital Epidemiology. https://doi.org/10.1086/668770
- World Health Organization. (2014). Antimicrobial Resistance: Global Report on Surveillance. WHO Report. https://doi.org/1.4.2014.